

嘉南藥理科技大學專題研究計畫成果報告

利用 PCR 檢測食品中 *Salmonella Enteritidis*

計畫類別：個別型計畫 整合型計畫

計畫編號：90 - FH - 02

執行期間：90年1月1日至90年12月31日

計畫主持人：王淑珍

執行單位：嘉南藥理科技大學 食品衛生系

摘 要

本實驗以 PCR(polymerase chain reaction) 檢測食品中 *Salmonella*, 選用的 primer 為 Sef127L-Sef661R)對 *Salmonella* Enteritidis 具有特異性, 其 PCR 產物為 535bp。由結果顯示對 27 株 *S. Enteritidis* 具有很高的專一性, 對於與 *Salmonella* 血緣相近及一些腸內菌, 皆無假性正反應產生。

緒 言

Salmonella 為引起人畜中毒之重要病原菌, 傳統檢測需經增菌、分離培養、純化、血清試驗及生化試驗, 需 6-7 天。進年來 polymerase chain reaction (PCR)技術以應用於病原菌之檢測(1,2)。對於 *S. Enteritidis* 之檢測, 有使用對 virulence plasmid 具特異性的 primer 來檢測 for *S. Enteritidis* (3,4)。以多套式 PCR(multiplex PCR) 來檢測 *S. Enteritidis*, 有檢測所有血清型 *Salmonella* 及 確認 *S. Enteritidis* 及 *S. Typhimurium*(5,6)。而本研究所使用的 PCR primer 與其他學者所發表之 PCR primers 不同。

PCR primer (Sef127L-Sef661R)為 *Salmonella* 鞭毛蛋白的一段基因, 對 *S. Enteritidis* 具有特異性。

材料與方法

1. PCR primer : Sef127L-Sef661R
2. DNA 製備: 培養後的菌液, 經煮沸作粗萃取液。
3. PCR 增幅作用: 94°C 50sec、60°C 50sec、72°C 40sec, 共 35cycles, 最後於 72°C 2min。
4. *Salmonella* 及相關細菌之檢測:
5. *Salmonella* Enteritidis 之靈敏度:
6. 南方吸漬法:

結果與討論

Salmonella Enteritidis 特異性:

由實驗結果顯示, primer Sef127L-Sef661RPCR 與 27 株 *Salmonella* Enteritidis 有正反應, 而其他非 *S. enteritidis* 之 *Salmonella* 及血緣相近之腸內菌皆無反應 (Table 1, Table 2)。為了確認 PCR 產物, 我們合成一段 checking probe 進行 Southern blot hybridization, 結果如 Fig 1 所示, 顯示 primer Sef127L-Sef661RPCR 增幅之 PCR 產正確性。

檢測靈敏度：

檢測結果顯示，經稀釋加熱萃取 DNA，每次只要 104CFU，就可檢測出來 (Fig2)。若要提高檢測靈敏度，可採預培養及選擇性培養來提高結果靈敏度。

參考文獻

1. Aderson, M.R. and C.J. Omiecinski. 1992. Direct extraction of bacterial plasmids from food for polymerase chain reaction amplification. *Appl. Environ. Microbiol.* 58:4080-4082.
2. Gouws, P.A., M. Visser and V.S. Brozel. 1998. A polymerase chain reaction procedure for the detection of *Salmonella* spp. Within 24 hours. *J. Food Prot.* 61:1093-1042.
3. Fadl, A.A., A.V. Nguyen and Khan, M.I. 1995. Analysis of *Salmonella enteritidis* isolates by arbitrarily primed PCR. *J. Microbiol.* 33:987-989
4. R.W. Moore, D.E. Corrier, L.H. Stanker, and B.M. Hargis. 1998. Comparison of Enrichment methods for recovery and chick infectivity of chlorine injured *Salmonella enteritidis*. *J. Food Prot.* 61:1054-1056.
5. Soumet, C.G., E.V. Rose, R.P. Drouin, G. Salvat and P. Colin. 1999. Identification by a multiplex PCR based assay of *Salmonella typhimurium* and *Salmonella enteritidis* strains from environmental swabs of poultry houses. *Letter Appl. Microbiol.* 29:1-6.
6. Cohen, N., E.D. Mcgruder, H.L. Neibergs, R.W. Behle, E.E. Wallis and B.M. Hargis. 1994. Detection of *Salmonella enteritidis* in feces from poultry using booster polymerase chain reaction and oligonucleotide primers specific for all members of the genus *Salmonella*. *Poultry Sci.* 73:354-357.

Table 1. PCR results for *Salmonella* isolates

Species	No. of isolates	positive results SefB127L-661R	Species	No. of isolates	Positive results SefB127L-661R
<i>S. aberdeen</i>	1	0	<i>S. kuru</i>	1	0
<i>S. adelaide</i>	1	0	<i>S. lagos</i>	1	0
<i>S. agona</i>	1	0	<i>S. lanka</i>	1	0
<i>S. albany</i>	1	0	<i>S. limete</i>	1	0
<i>S. amager</i>	1	0	<i>S. litchfield</i>	1	0
<i>S. anatum</i>	2	0	<i>S. london</i>	1	0
<i>S. allandale</i>	1	0	<i>S. meleagridis</i>	1	0
<i>S. azteca</i>	1	0	<i>S. manhattan</i>	1	0
<i>S. bareilly</i>	1	0	<i>S. minnesota</i>	1	0
<i>S. berta</i>	3	0	<i>S. miami</i>	1	0
<i>S. bonn</i>	1	0	<i>S. montevideo</i>	2	0
<i>S. boussol</i>	1	0	<i>S. muenchen</i>	1	0
<i>S.</i>	1	0	<i>S. munster</i>	2	0
<i>bovismorbificans</i>					
<i>S. bredeny</i>	1	0	<i>S. nchanga</i>	1	0
<i>S. braenderup</i>	2	0	<i>S. newbrunswick</i>	1	0
<i>S. cairo</i>	1	0	<i>S. newington</i>	1	0
<i>S. californica</i>	1	0	<i>S. newport</i>	2	0
<i>S. cerro</i>	1	0	<i>S. ngor</i>	1	0
<i>S. chailey</i>	1	0	<i>S. nigeria</i>	1	0
<i>S. chester</i>	1	0	<i>S. ohio</i>	1	0
<i>S. colorado</i>	1	0	<i>S. oranienburg</i>	1	0
<i>S. choleraesuis</i>	1	0	<i>S. panama</i>	2	0
<i>S. coleypark</i>	1	0	<i>S. portsmouth</i>	1	0
<i>S. cubana</i>	1	0	<i>S. richmond</i>	1	0
<i>S. derby</i>	2	0	<i>S. rubislaw</i>	1	0
<i>S. djakarta</i>	1	0	<i>S. sandiego</i>	1	0
<i>S. drypool</i>	1	0	<i>S. senftenberg</i>	3	0
<i>S. enteritidis</i>	27	27	<i>S. seremban</i>	1	0
<i>S. eppendorf</i>	1	0	<i>S.</i>	1	0
			<i>schwarzengrund</i>		
<i>S. essen</i>	1	0	<i>S. sinstorf</i>	1	0
<i>S. gera</i>	1	0	<i>S. stanley</i>	1	0
<i>S. goerlitz</i>	1	0	<i>S. tananarive</i>	1	0
<i>S. hadar</i>	3	0	<i>S. tennessee</i>	2	0
<i>S. halmstad</i>	1	0	<i>S. thomasville</i>	1	0
<i>S. hartford</i>	1	0	<i>S. thompson</i>	1	0
<i>S. havana</i>	2	0	<i>S. trachau</i>	1	0
<i>S. heidelbrg</i>	1	0	<i>S. typhimurium</i>	4	0
<i>S. hvittingfoss</i>	1	0	<i>S. vejle</i>	1	0
<i>S. infantis</i>	3	0	<i>S. victoria</i>	1	0
<i>S. johannesburg</i>	2	0	<i>S. weltevreden</i>	1	0
<i>S. kentucky</i>	1	0	<i>S. worthington</i>	1	0
<i>S. kinshasa</i>	1	0			

Table2. PCR results for non-*Salmonella* isolates

Species	No. of Isolates	positive	results
		SefB 127L-SefB	661R
<i>Escherichia coli</i>	23		0
<i>Escherichia coli (ETEC)</i>	9		0
<i>Staphylococcus aureus</i>	7		0
<i>Acinetobacter calcoaceticus</i>	1		0
<i>Alcaligenes faecalis</i>	1		0
<i>Bacillus stearothermophilus</i>	1		0
<i>Bacillus subtilis</i>	1		0
<i>Brevibacterium linens</i>	1		0
<i>Citrobacter freundii</i>	2		0
<i>Enterobacter aerogenes</i>	2		0
<i>Enterobacter cloacae</i>	1		0
<i>Erwinia carotovora</i>	1		0
<i>Hafnia alvei</i>	1		0
<i>Proteus vulgaris</i>	2		0
<i>Salmonella arizonae</i>	1		0
<i>Serratia marcescens</i>	1		0
<i>Shigella flexneri</i>	1		0
<i>Shigella sonnei</i>	1		0

ATCC: American Type Culture Collection, Maryland, U.S.A.

CDC: Center for Disease Control, Georgia, U.S.A.

PT : National Ping Tung Institute of Agriculture.

US: The City of New York Department of Health. U. S. A.

CCRC: Culture Collection and Research Center Taiwan R.O.C.

WHO: World Health of Organization.

FDB: Food Drug Bureau, Department of Health Executive Yuan, Taiwan. R. O. C.

VSE: Strains of *S. enteritidis* from Veterinary Service Laboratory of USDA.

ISE: Strains of *S. enteritidis* from National Institute of preventive Medicine, Department of health, executive Yuan, Taipei, Taiwan, R.O.C.

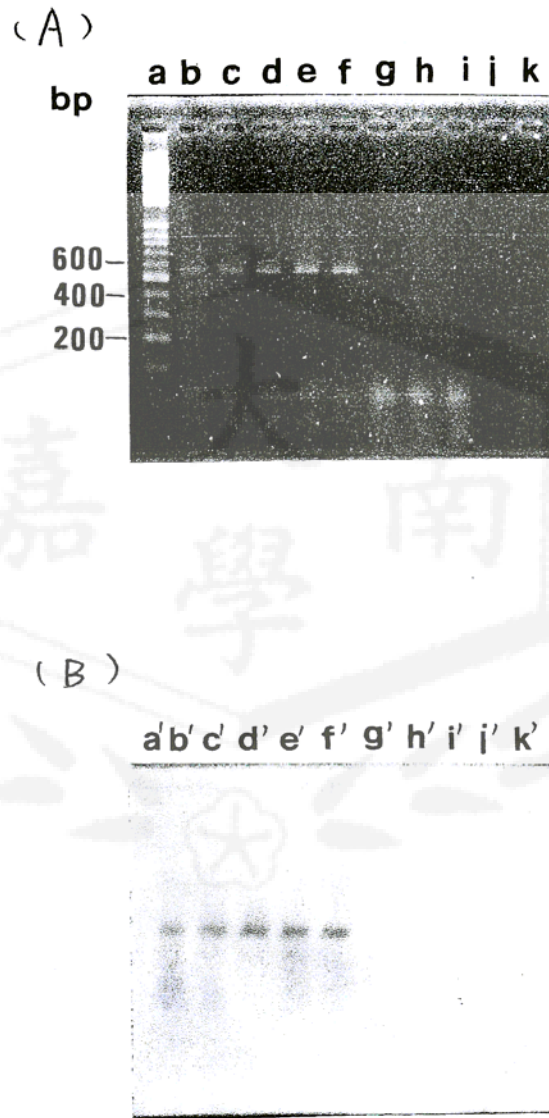


Fig.1 Agarose gel electrophoresis of the PCR products amplified from *Salmonella enteritidis* with PCR primer SefB127L-661R (A) southern blot hybridization of the PCR products with digoxigenin-labeled oligonucleotide (B) Lane a :marker ; b~f : *Salmonella enteritidis* ; g : *Salmonella anatum* ; h : *Salmonella raenderup* ; i : *Escherichia coli* ; j : *Escherichia coli*(EIEC03) ; k : *Bacillus subtilis* .Lane b'~k': southern blot hybridization results for Lane b~k in Fig.1(A)

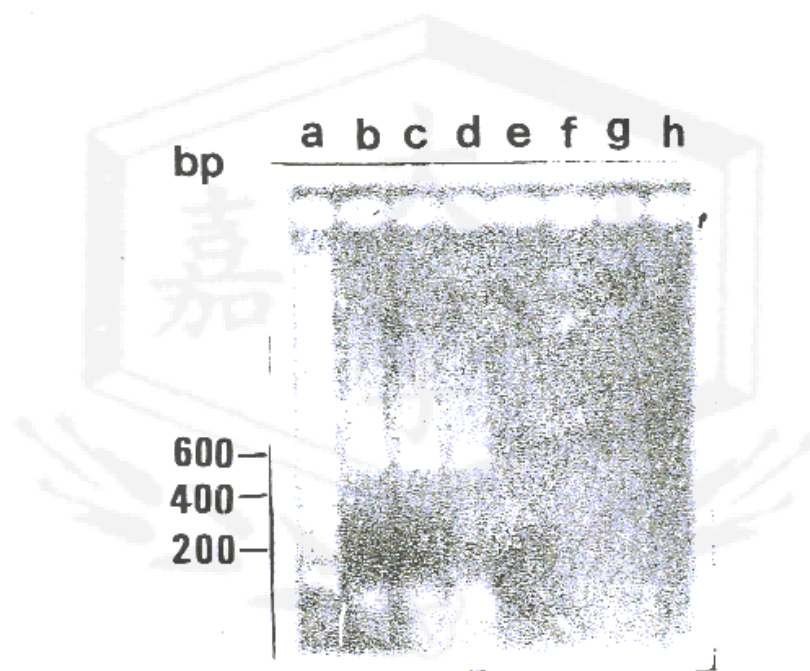


Fig.2 Detection sensitivity for *Salmonella enteritidis* using PCR primer SefB127L-661R , Lane a : marker ; b~d : *Salmonella enteritidis* were spiked 10^9 , 10^8 , 10^7 CFU/ml respectively.